

# Computer implementation of myogenic gradient in the retinal network

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## Abstract:

Vascular network in the retina is a good object for in vivo investigation of the blood flow. Different techniques are used for deep study of blood flow and its characteristics, including computational methods. We suggest quantitative mathematical model of retinal vascular network, that represent basic features of the system.

## 1. Why the retina?

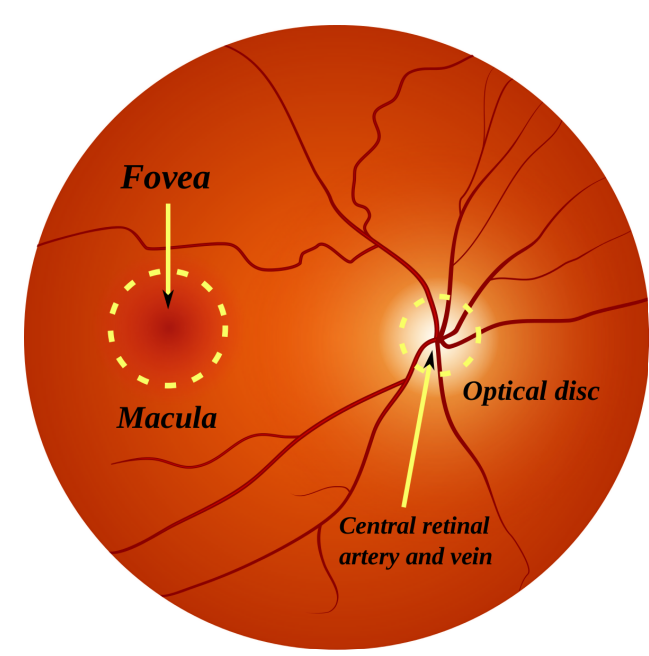


Figure 1: Typical retinal image as seen through the pupil

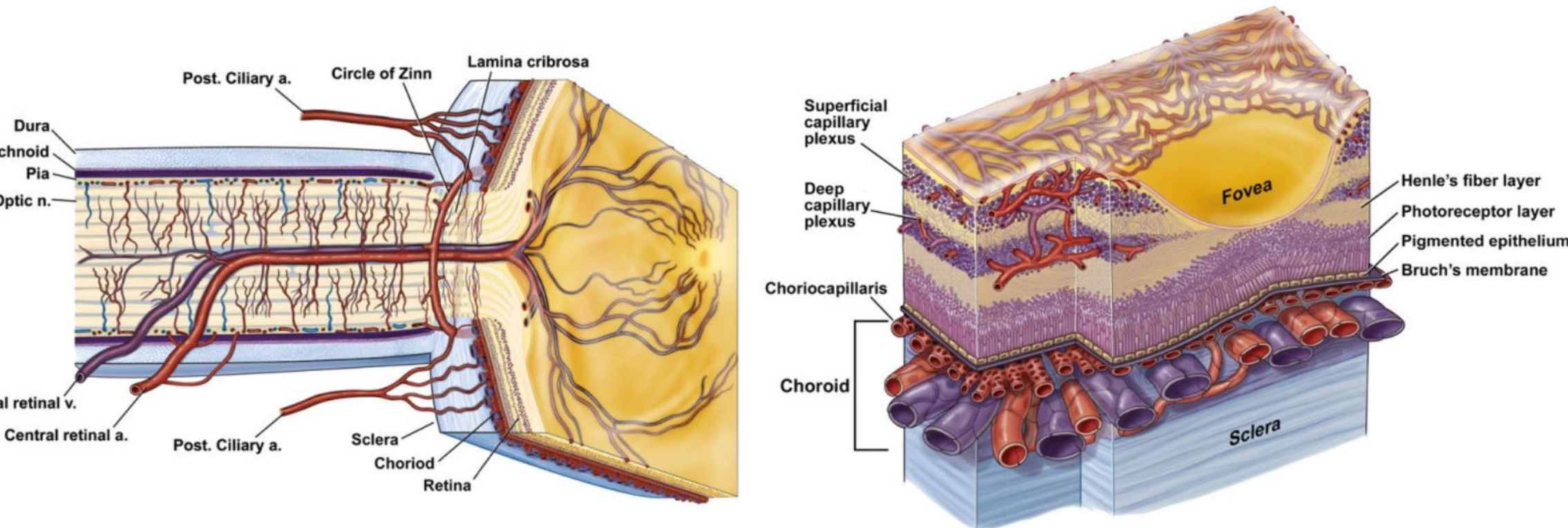


Figure 2: Anatomy of ocular circulation (a-artery, b-vein, n-nerve) a) Cut away drawing along the superior-inferior axis of the human eye through the optic nerve, showing the vascular supply to the retina and choroid. b) Drawing showing vasculature of the retina and choroid. Drawings by Dave Schumick from Anand-Apte and Hollyfield (2009).

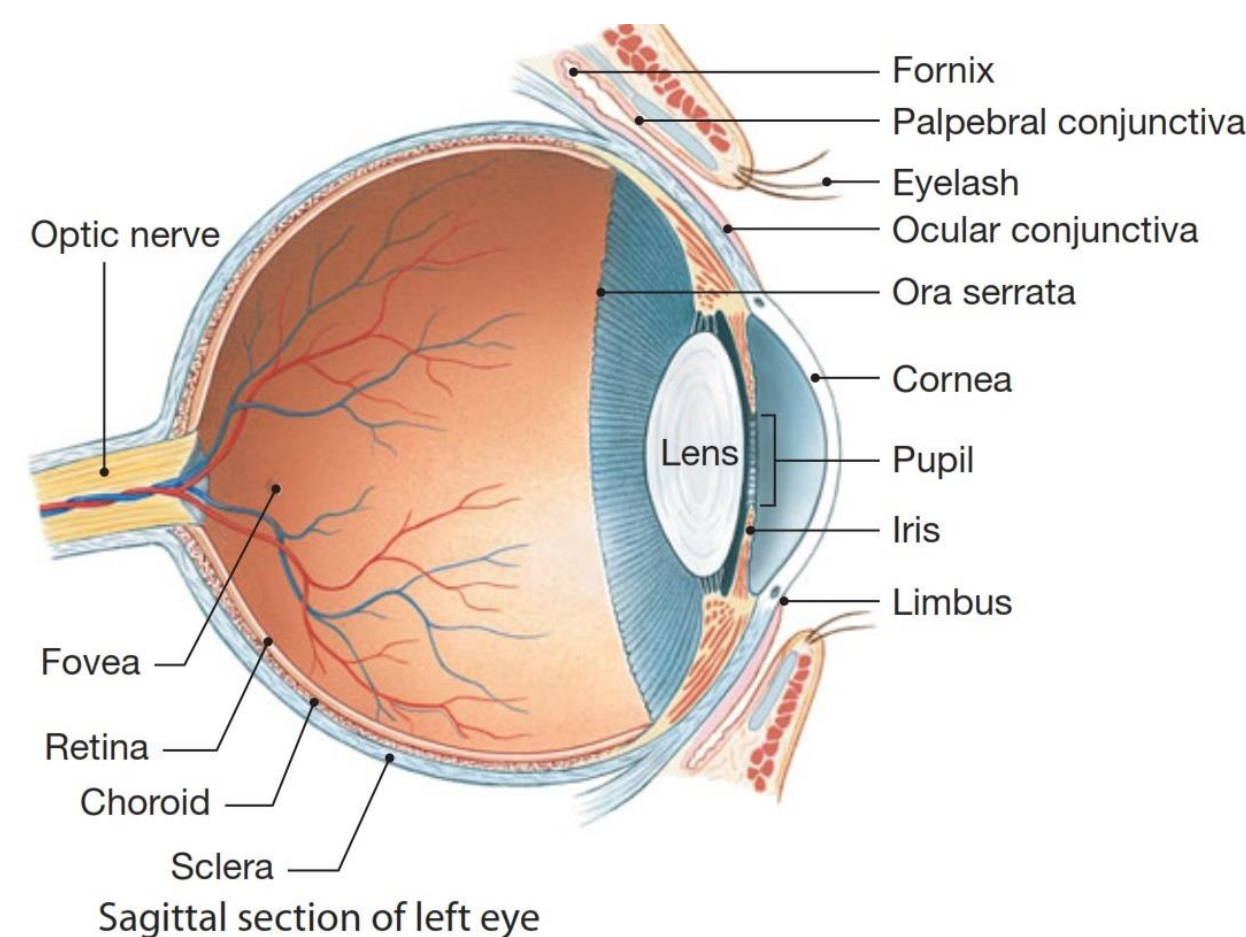


Figure 3: Sagittal section of left eye. Drawing from Martini (2011).

## 2. Modelling of the retinal vessel network

### 2.1 Network structure

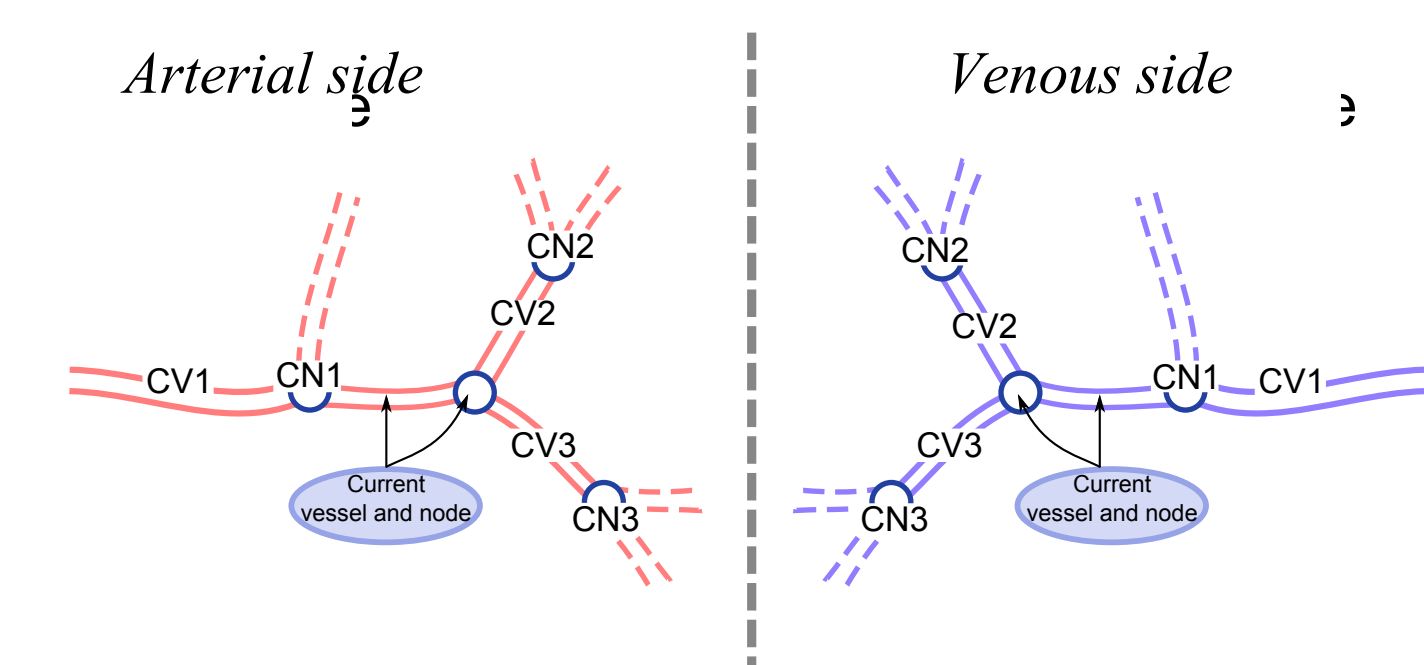


Figure 4: Schematic representation of vessel network. For each vessel on the each side (arterial or venous) we have 1 parent vessel CV1 and 1 parent node CN1 and 2 daughter vessels (CV2 and CV3) and daughter nodes (CN2 and CN3). These denotations are used for making of Vessel Matrix and Node Matrix

To find vessels lengths and radiuses we used relations:

$$r_p^3 = r_{d1}^3 + r_{d2}^3 + r_{d3}^3 + \dots + r_{dn}^3 \quad \text{Murray's law, where } r_p - \text{radius of parent vessel } r_d - \text{radius of daughter vessel}$$

$$h = 0.14r_i + 3.3, \quad \text{where } h - \text{vessel wall thickness, } r_i - \text{vessel inner radius (Hashimoto, 1986)}$$

The pressure in each nodes calculates using Kirchhoff's law:

$$\sum_j Q_j^n = \sum_j C_j^n \Delta P_j^n = 0, \quad \text{where is the } Q_j^n - \text{inlet/outlet flow, } C_j^n - \text{the vascular conductance of the } j^{\text{th}} \text{ vessel and } \Delta P_j^n - \text{pressure drop in the related vessel.}$$

### 2.2 Vessel walls' mechanics

Vessel wall stress relations:

$$\sigma = \sigma_p \cdot af_p + \sigma_a \cdot af_a + \sigma_c \cdot af_c$$

$$\sigma_p = C_{p1} \cdot (e^{a_{p1} \cdot \epsilon} - 1) + C_{p2} \cdot (e^{a_{p2} \cdot \epsilon} - 1)$$

$$\epsilon = \frac{r_m - r_{slack}}{r_{slack}}$$

$$\dot{\epsilon} = k_1 \cdot (\bar{\sigma} - \sigma) \cdot \left( \frac{l - l_{aopt}}{b_a} \right)^2$$

$$\sigma_{am} = \sigma_{amopt} \cdot e^{-\left( \frac{l - l_{aopt}}{b_a} \right)^2}$$

$$\sigma_a = \psi \cdot \sigma_{am}$$

$$\sigma_c = \sigma_{c0} \cdot \frac{e^{b_c \cdot (l - l_{c0})} - e^{-b_c}}{1 - e^{-b_c}}$$

$$\bar{\sigma} = P \cdot r_i / h$$

Vessel tone relations:

$$\dot{\psi} = k_2 (\bar{\psi} - \psi)$$

$$\bar{\psi} = \left( \frac{(\sigma + CON)^{hc}}{(\sigma + CON)^{hc} + \sigma_{50}^{hc}} \right)$$

$\sigma$  - mean wall stress;  
 $\sigma_p, \sigma_a, \sigma_c$  - stress on element type  $j, j = p(\text{active}), a(\text{active})$  or  $c(\text{cytoskeleton})$ ;  
 $af_p, af_a, af_c$  - fraction of wall area covered by matrix, active/cytoskeletal elements;  
 $C_{p1}, C_{p2}$  - constants in passive stress-strain curve;  
 $a_{p1}, a_{p2}$  - constants in passive stress-strain curve;  
 $\epsilon$  - strain;  
 $r_m$  - midwall radius;  $r_{slack}$  - slack radius (radius at zero load);  
 $k_1$  - rate constant mechanical radius changes;  
 $\sigma_{am}$  - capacity for act. stress generation;  $\sigma_{amopt}$  - max. act. stress at optimal length;  
 $l$  - SMC length;  $l_{aopt}$  - optimal SMC length for active stress;  
 $b_a$  - width of active length-stress curve;  
 $\psi$  - vessel tone;  
 $\sigma_{c0}$  - cytoskeletal stress at  $l_{c0}$ ;  $b_c$  - steepness cytoskeletal stress curve;  
 $P$  - midvessel pressure;  $r_i$  - inner radius;  $h$  - wall thickness;

$CON$  - constrictor;  $hc$  - Hill coefficient for act. curve;  
 $\sigma_{50}$  - stress for half-maximal activation;  
 $k_2$  - rate constant tone changes;

Ref.: Integrative Modeling of Small Artery Structure and Function Uncovers Critical Parameters for Diameter Regulation, Ed Van Bavel (2014)

### 2.3. Modelling of myogenic gradient

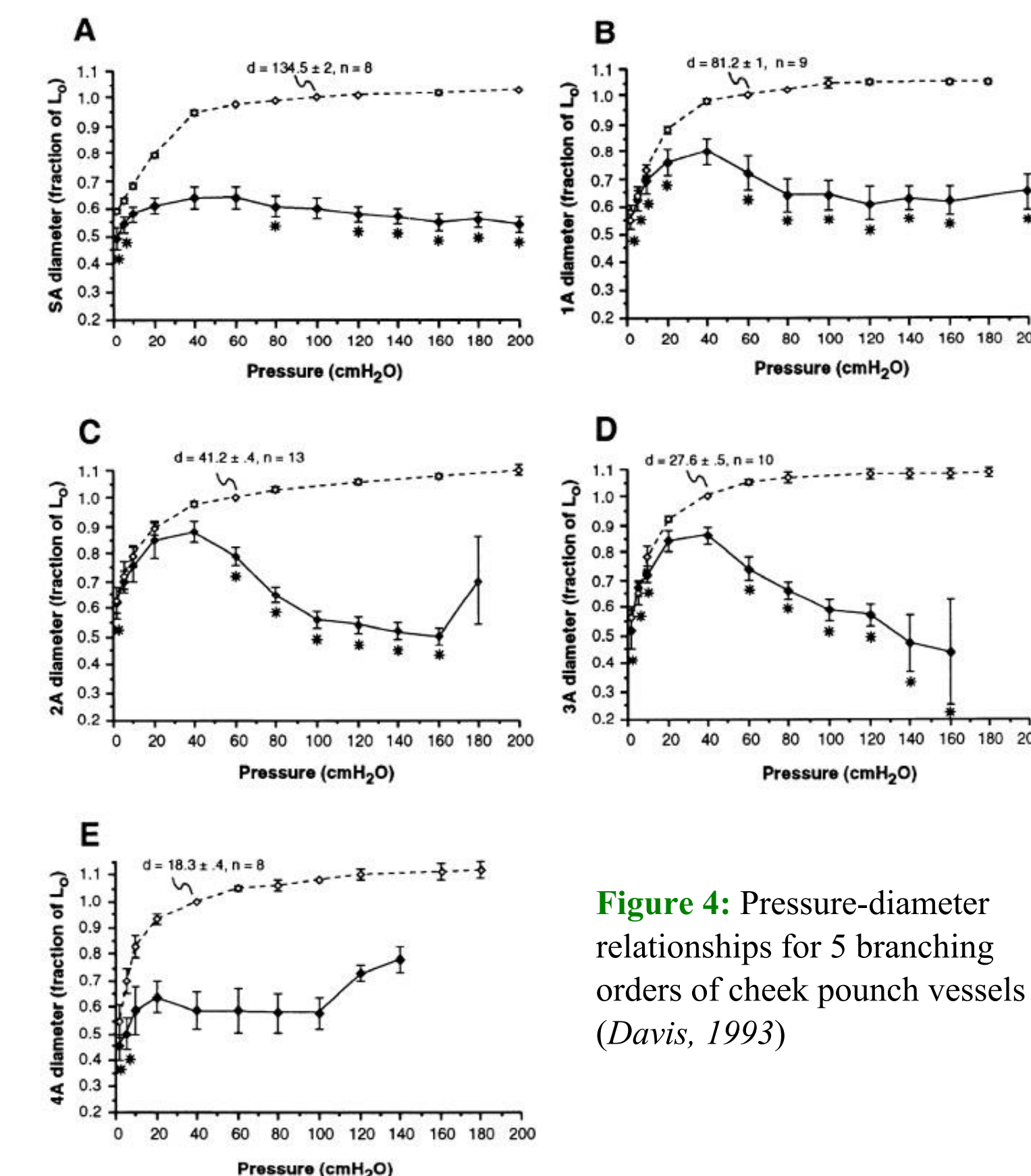


Figure 4: Pressure-diameter relationships for 5 branching orders of cheek pouch vessels (Davis, 1993)

• Myogenic response is reaction of smooth muscle in the vessel wall on the external stimulus, that contacts in response to increasing intraluminal pressure and relax in response to decreasing pressure, in order to balances optimal vessel diameter.

• Myogenic response observed in different vascular beds in both small and large vessels.

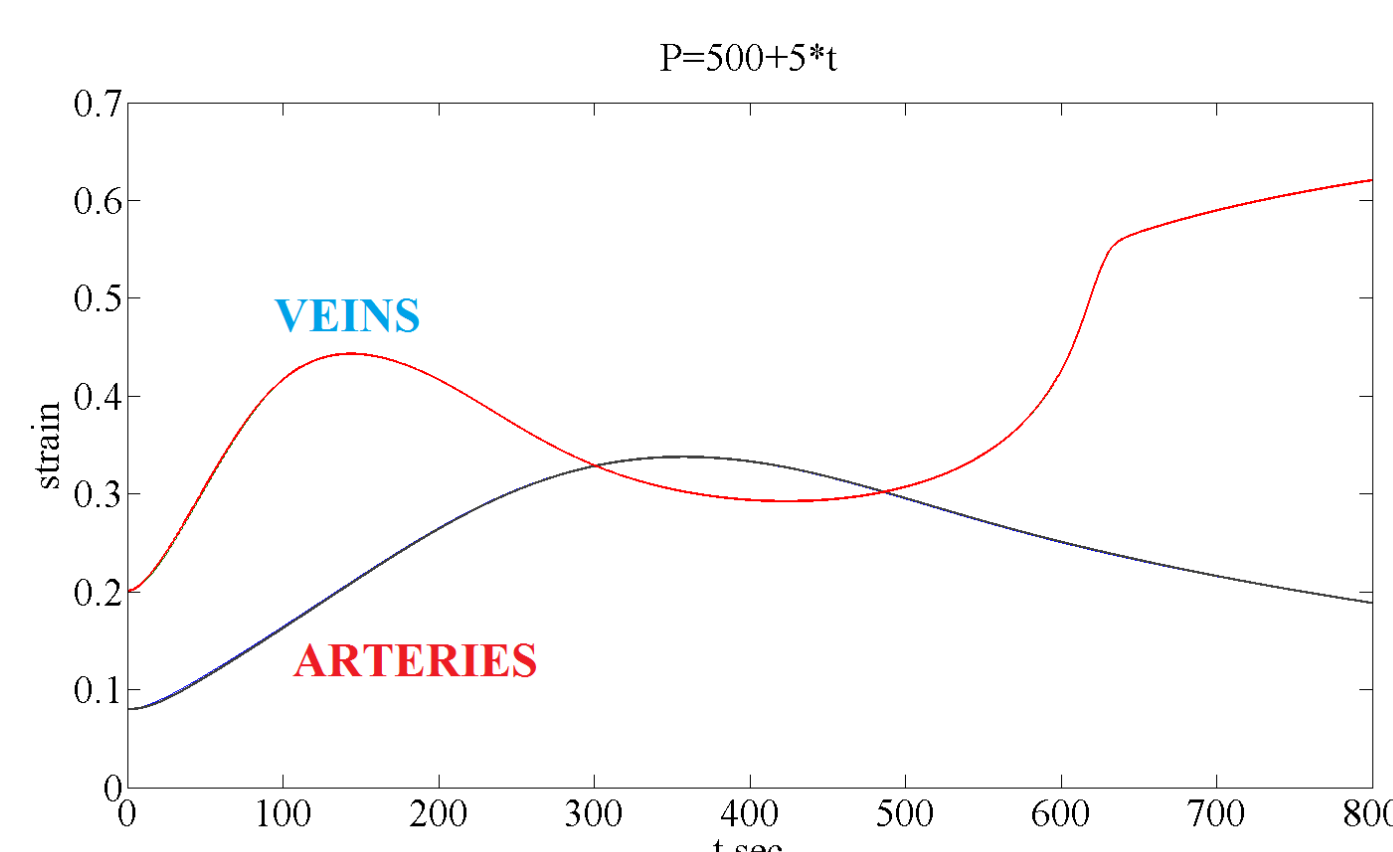
• It was shown that, intensity of myogenic response of hamsters' isolated arterioles depends from vessel radius and most intensive for the terminal vessels (Davis, 1993).

• Also, it was found, that myogenic response gradient are also exist in human retinal arterioles (Jeppesen et al., 2007).

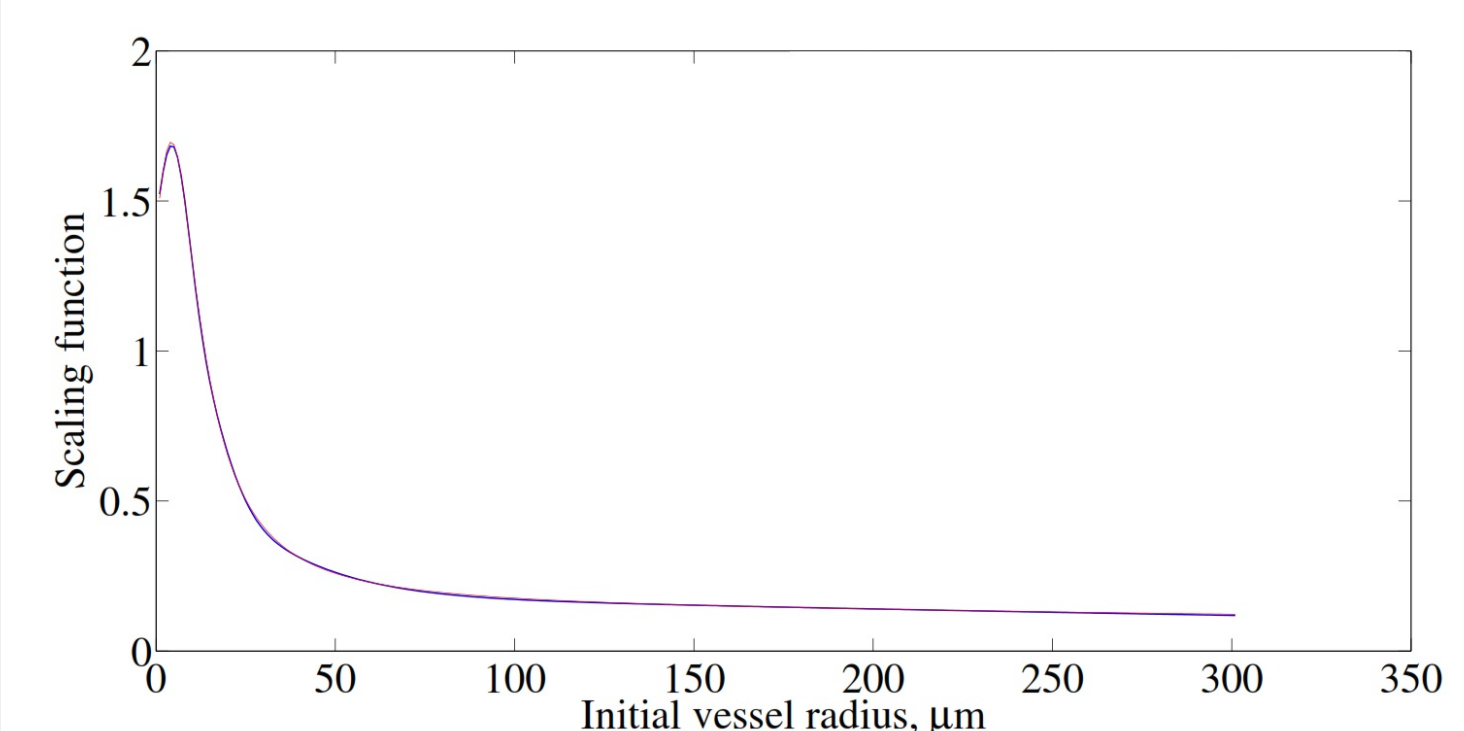
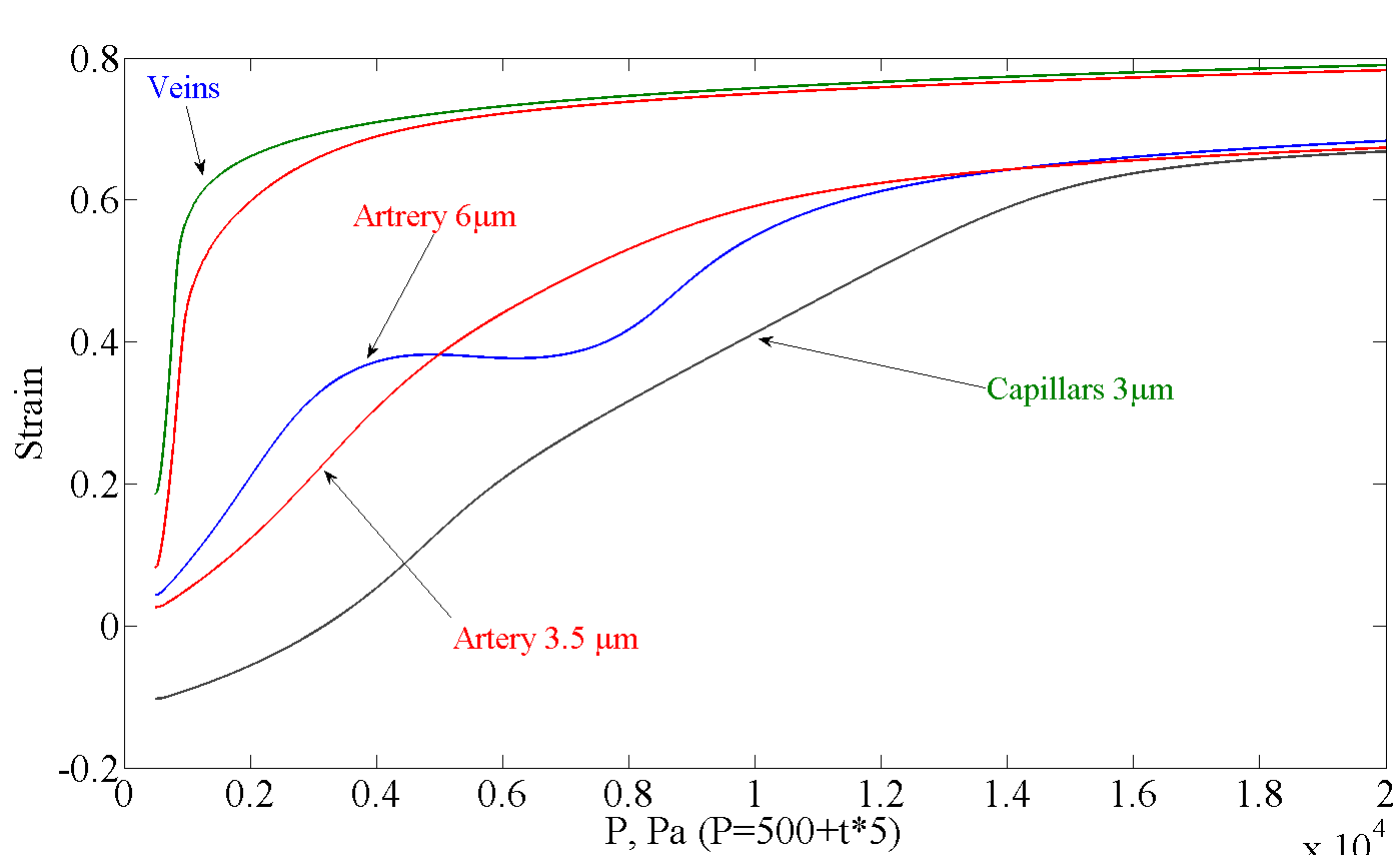
• Longevity of myogenic response depends from vessel radius, too. Vessels with smaller diameter, and, hence, thicker vessel wall are able to maintain smaller vessel radius during increasing inlet pressure less than larger vessels.

To take into account these features of myogenic response we introduces function of scaled mean stress

Myogenic response in the retinal network without dependence from initial vessel radius



Myogenic response in the retinal network with dependence from initial vessel radius



We introduced new variable - scaled mean stress  $\sigma_{sc}$  which reflects dependence of mean stress (where  $h$  - wall thickness) from initial vessel radius:

$$\sigma = P \cdot r_i / h$$

$$\sigma_{sc} = \sigma \cdot \text{Scalefunc}$$

$$\text{Scalefunc} = a_1 e^{-((radius-b_1)/c_1)^2} + a_2 e^{-((radius-b_2)/c_2)^2} + a_3 e^{-((radius-b_3)/c_3)^2} + a_4 e^{-((radius-b_4)/c_4)^2},$$

where:

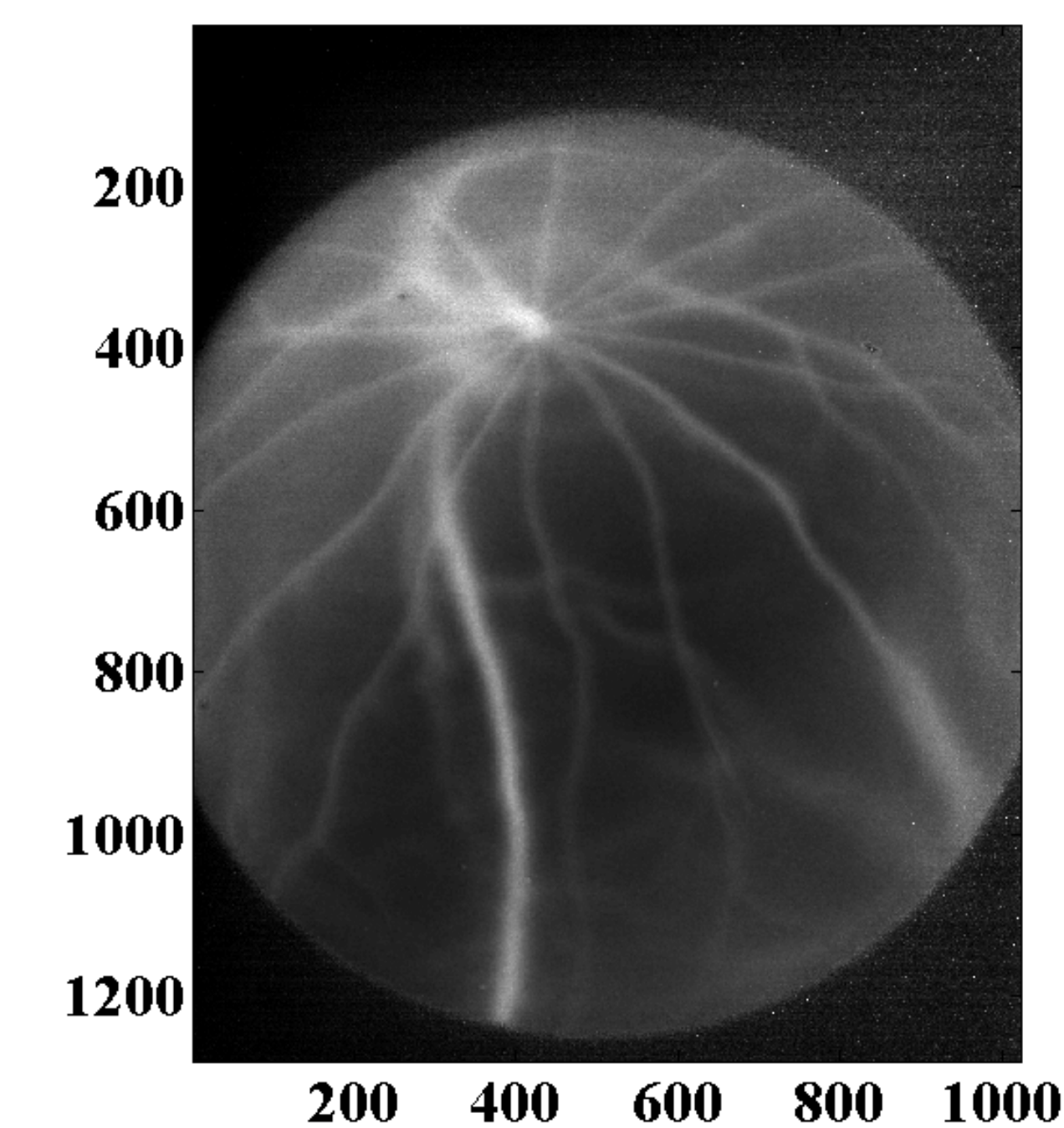
$$a_1 = 0.6357, b_1 = 4.336, c_1 = 6.62$$

$$a_2 = 0.4003, b_2 = 8.791, c_2 = 12.59$$

$$a_3 = 4.785, b_3 = -128.9, c_3 = 88.75$$

$$a_4 = 1.975e+04, b_4 = -1.402e+04, c_4 = 4130$$

### Average blood flow image



Experimental setup includes:

- Endoscope
- Laser
- Camera
- PC software

Future plans:

- LSI experiment, tests with chemicals which induce constriction/relaxation retinal vessel
- Rewriting mathematical model on CUDA C language
- Test simulations on realistic network with thousands vessels, find the circumstances under which network could show vasomotion

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### Rat retina Laser Speckle Imaging (Blood flow visualization)

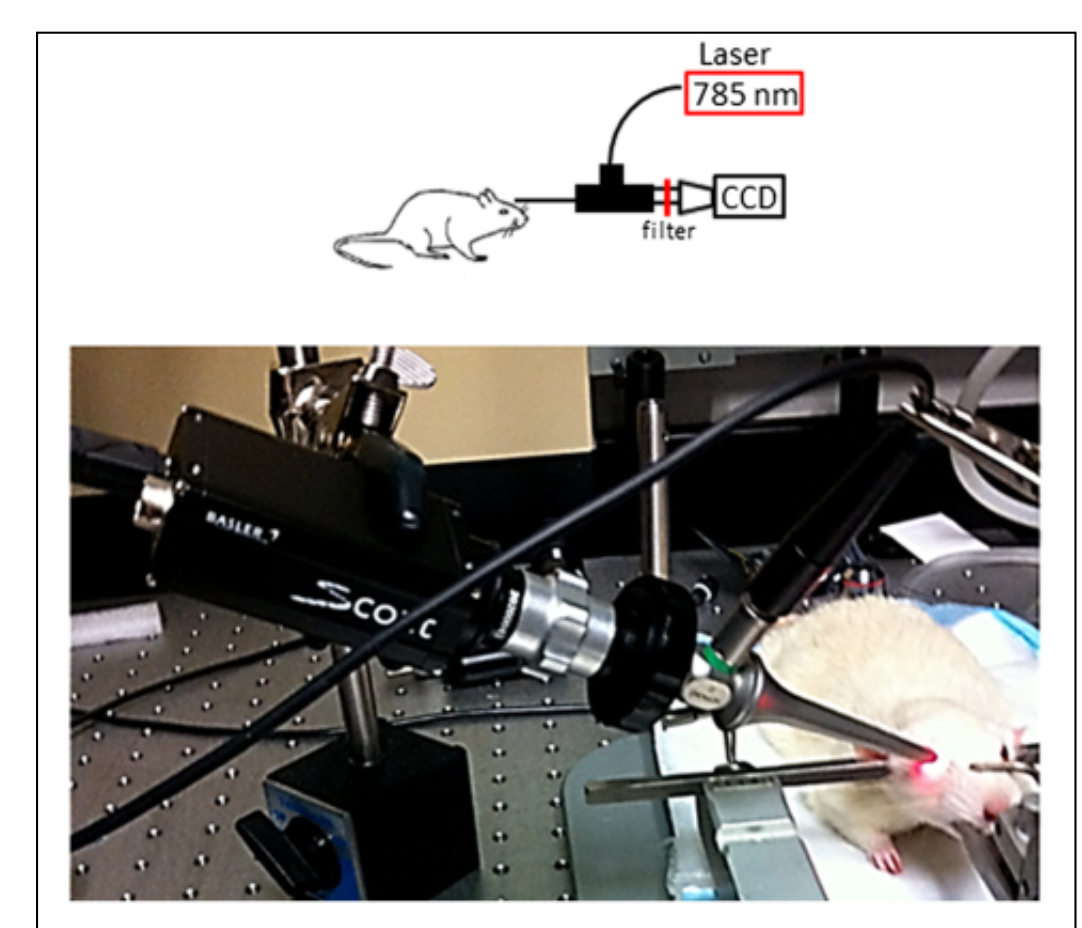


Figure 9: Schematic of imaging setup. A photograph of the imaging system is also shown (Drawing from Ponticorvo, 2013)

### Blood flow image

